

The adipose tissue as a source of vasoactive hormones and cytokines with a potential role in the pathogenesis of cardiovascular and renal diseases

Jerzy Chudek, Marcin Adamczak, Teresa Nieszporek, Andrzej Więcek

Department of Nephrology, Endocrinology and Metabolic Diseases.
Medical University of Silesia, Katowice, Poland

SUMMARY

During the last decade white adipose tissue has been recognised as an active endocrine organ and a source of many hormones and proinflammatory cytokines in obesity. These adipokines may play an important role in the pathogenesis of cardio-vascular and kidney diseases.

The contribution to the vascular pathology of obesity of different cell types which compose the adipose tissue; adipocytes, preadipocytes, stromal/vascular cells and macrophages, is, however, different.

In this review we have summarized the present knowledge of some adipokines in patients with obesity, arterial hypertension and chronic kidney diseases.

Key words: leptin, adiponectin, resistin, interleukin-6, TNF-alpha, cardio-vascular.

INTRODUCTION

The epidemic of visceral obesity, insulin resistance, noninsulin-dependent diabetes mellitus and obesity related arterial hypertension is a challenging health problem for modern societies. No solution to the increasing danger posed by these well known risk factors of cardiovascular morbidity and mortality has been yet pro-

Received for publication: 24/08/2005

Accepted: 14/09/2005

posed. Propagation of an active life style and low caloric diet as well as anorexigenic medications are still insufficient to counterbalance an easy approach to high caloric products.

The last decade has seen adipose tissue re-discovered as an active endocrine organ and an important source of several proinflammatory cytokines^{1,2}. Several of these may directly influence the function of the cardio-vascular system and the atherosclerosis process. An incomplete list of adipokines would run as follows: leptin, adiponectin and resistin, estrogens and glucocorticoids, renin, angiotensin II, PAI-1, tumour necrosis factor- α (TNF- α), interleukin-6, interleukin-8, interleukin-10, interleukin-1 β , acylation stimulating protein (ASP), prostaglandin E₂ (PGE₂), hepatocyte growth factor (HGF), vascular endothelial growth factor (VEGF), insulin growth factor-1 (IGF-1), tissue factor (TF), complement factor D (adipsin), Agouti signaling protein (ASP), nitric oxide (NO)³.

Adipose tissue is not a homogenous organ. It consists of a variety of different cell types such as adipocytes, preadipocytes, stromal/vascular cells and macrophages³ (Table I). Each of these cells present their own secretion profile and specific regulation. It is already well known that mature adipocytes are the main source of leptin and adiponectin, that macrophages produce almost all TNF- α , while PGE₂, interleukins, VEGF, HGF are synthesized by stromal and vascular cells^{3,4}. There are also some differences in the adipokines production between visceral and peripheral fat tissue (Table II). Adipokines released from the fat tissue may exert their action on the endocrine, paracrine or autocrine pattern. While this review mainly focuses on a discussion of the endocrine action of adipokines, the paracrine action can not be neglected in cardio-vascular diseases. An example of this is secretion of NO by the periadventitial fat tissue⁵. Its action on the neighbouring smooth muscle cells may actively

influence the vascular tonus and arterial blood pressure⁵.

The knowledge of adiposity related mechanisms devastating the cardio-vascular system and the kidneys may help in the design of new drugs for the prevention and treatment of the metabolic syndrome. The role of some adipokines in the pathogenesis of cardio-vascular and renal diseases is discussed in this review.

Leptin

Leptin is a protein predominantly produced by adipocytes⁶. It is encoded by the *ob* gene⁶. Initially, leptin was implicated in the regulation of appetite as a satiety hormone⁶. It was found that rats with homozygous nonsense mutation of the *ob* gene were suffering from marked obesity, while parenteral leptin substitution was decreasing their appetite and body mass⁷. A few years later leptin appeared mostly as a marker of nutrition unable to decrease food intake in obese humans, as opposed to what was initially expected. Obese individuals, especially females, are characterised by high plasma leptin concentrations⁸. Thus even highly elevated plasma leptin concentration does not suppress the appetite in these individuals⁹.

According to Flier¹⁰, leptin should not be regarded as an antiobesity hormone but as a sign of energy deficiency and integrator of neuroendocrine function. Low plasma leptin concentration – a 'starvation signal' – modulates the hypothalamic-pituitary-adrenal axis, suppresses thyroid and gonadal axes¹⁰. These endocrine changes resemble the clinical status of patients with *anorexia nervosa*¹¹.

Low plasma leptin concentration decreases energy expenditure and stimulates the search for food in deprived of food rodents^{12,13}. Only in animals with low body fat stores it was demon-

TABLE I
The most important adipokines released by adipocytes and the matrix of adipose tissue

	Adipocytes	Tissue matrix
Prostaglandin E ₂	+/-	+++
Prostacycline	+/-	+++
Adiponectin	+++	+
Leptin	+++	-
Resistin	+	+++
Interleukin 8	+	+++
Interleukin 6	+/-	+++
Interleukin 10	+/-	+++
Interleukin 1β	+/-	+++
Tumor necrosis factor α (TNF-α)	+/-	+++
Platelet activator inhibitor 1 (PAI-1)	++	+++
Hepatocyte growth factor (HGF)	+/-	+++
Vascular epithelial growth factor (VEFG)	+/-	+++
Angiotensin II	+	+++

TABLE II
Comparison of gene expression of adipokines by adipocytes in human visceral or subcutaneous adipose tissue

	Visceral	Subcutaneous
Leptin	+	++
Adiponectin	+++	+
IL-6	+++	+
TNF-α	+	+
PAI-1	++	+
Angiotensinogen	++	+
Estrogens	+	+
IGF-1	+	+

trated that leptin is involved in the regulation of food intake and energy expenditure. Similarly, only in obese children with inactivating mutations of both leptin alleles, parenteral substitution of this peptide decreases appetite with a subsequent reduction of body mass¹⁴.

It is well known that sexual maturation and fertility are linked to nutritional status and adiposity. Injection of leptin in the prepubertal female mouse causes earlier maturation of the reproductive tract¹⁵, suggesting that leptin acts as a signal for puberty. Moreover leptin may promote angiogenesis and stimulate proliferation and differentiation of haemopoietic cells, including T cells^{16,17}. In acute infections the shift of the immune system, stimulated by leptin, toward the predominance of the proinflammatory Th₁ T cells population seems to be beneficial¹⁸. Chronic stimulation of the immune system by leptin may lead to acceleration of atherosclerosis, as leptin-deficient mice are protected from atherosclerosis despite all the other metabolic factors that contribute to this vascular disease¹⁹.

In obese humans leptin exerts a deteriorative impact on the cardiovascular system and kidneys by significant contribution to the pathogenesis of obesity related arterial hypertension. It was shown that high plasma leptin concentration stimulates the activity of the sympathetic nervous system via the receptors located in the brain trunk²⁰. Moreover, leptin exaggerates insulin resistance in obese patients²¹, stimulates activity of the renin-angiotensin system²² and modulates the function of endothelial cell (vascular remodelling)¹⁶. Many of these above mentioned mechanisms interfere with the tubular sodium reabsorption. The contra-regulatory mechanisms responsible for the lack of arterial hypertension in 10-20 % of obese humans were not identified.

Leptin is also involved in the pathogenesis of obesity-related nephropathy and the progression

of chronic glomerulopathies²³. Within the glomerulus, leptin directly stimulates endocapillary cell proliferation and mesangial collagen type I and IV deposition^{24,25}. In cultured rat endothelial cells, mouse recombinant leptin stimulates proliferation and increases TGF- β mRNA and TGF- β secretion²⁴. It also stimulates the expression of TGF receptors²⁴. Leptin infusion in rats enhances urinary protein excretion²⁴. In addition, leptin induces overactivity of the sympathetic nervous, that may contribute to renal damage directly or indirectly via elevated blood pressure.

Patients with advanced chronic kidney disease (CKD) are characterized by markedly elevated plasma leptin concentration compared to body mass index and sex matched healthy individuals²⁶⁻²⁸. CAPD patient have even higher plasma leptin concentration than haemodialysis patients^{26,27}. However, increased plasma leptin concentration in CKD is not accounted for by oversecretion of this hormone. Leptin gene expression is even lower in adipocytes of CKD patients than in healthy individuals²⁹. In renal failure decreased renal clearance of leptin contributes to elevated plasma leptin concentrations³⁰. Thus it is not surprising that plasma leptin concentration is normalized by successful kidney transplantation³¹.

Elimination of leptin is partly independent from glomerular filtration rate^{32,33}. Kinetic studies suggest that the active uptake of the peptide hormone by the renal tissue contributes to leptin catabolism³⁴. Cumin *et al.*³² studied plasma leptin in Zucker obese rats subjected to bilateral nephrectomy or bilateral ureteral ligation. Following the bilateral nephrectomy plasma leptin concentrations increased by 300%, more than the 50% increase observed after bilateral ligation of the ureters. Similarly in patients with noninflammatory, reversible acute renal failure requiring haemodialysis treatment, plasma leptin concentration although significantly higher than in

healthy subjects was much lower than in CKD patients on haemodialysis³³.

In chronic uremia elevated plasma leptin concentration seems to participate in the pathogenesis of uremia-associated cachexia via signaling through the central melanocortin system³⁵. On the other hand, it should be stressed that studies addressing the influence of plasma leptin concentration on future changes of the body mass in hemodialysis patients failed to find such a relationship^{36,37}.

Plasma leptin concentration may serve in these patients also as a marker of well being^{36,38}. Elucidation of the mechanisms regulating appetite will be useful in our understanding of cachexia in uremic patients. From the other side it is well known that low body mass predominantly muscle mass is combined with the increased morbidity and mortality in these patients. Therefore there is no doubt that these new findings on the role of leptin in the pathogenesis of cachexia in uremia may have important clinical and therapeutic implications³⁵.

Adiponectin

Adiponectin is the cardio protective protein hormone secreted almost exclusively by adipocytes with antiatherogenic and insulin-sensitizing properties. It shows structural homology with collagen VIII, X and complement factor C1q³⁹. Plasma concentration of adiponectin is relatively high (almost 0.01% of total plasma protein)⁴⁰. In contrast to leptin its concentration is lower in obese than in non-obese subjects⁴⁰. Lower adiponectinaemia was also found in males than in females, and in patients with coronary artery disease, diabetes mellitus type II and essential hypertension than in healthy subjects⁴¹⁻⁴³.

Adiponectin receptors are found in skeletal muscle, liver and endothelial cells⁴⁴. It has been

suggested that adiponectin inhibits formation of initial atherosclerotic lesions by decreasing expression of adhesion molecules (VCAM-1; ICAM-1, E-selectin) in endothelial cells in response to inflammatory stimuli such as $\text{TNF}\alpha$ ⁴⁵. Adiponectin also suppresses production of cytokines, such as $\text{TNF}\alpha$ by macrophages⁴⁶. Moreover, adiponectin suppresses accumulation of lipids in human monocyte-derived macrophages and inhibits transformation of macrophages into foam cells⁴⁷. It also inhibits cell proliferation stimulated by oxidized LDL and suppresses superoxide generation⁴⁸. Finally, adiponectin interferes with atherogenesis in ApoE-deficient mice, a well known model of spontaneous atherosclerosis⁴⁹.

Adiponectin participates also in the stabilization of atherosclerotic plaques by increasing expression of tissue inhibitor of metalloproteinase-1 (TIMP-1) in infiltrating macrophages⁵⁰.

Adiponectin improves insulin sensitivity⁴¹ by stimulating glucose utilization and fatty acid oxidation in skeletal muscles and the liver and by suppressing enhanced glucose production⁵¹. It has already been shown that low plasma adiponectin concentration is an independent predictor of the risk of developing insulin resistance and type 2 diabetes in the general population⁵². Adiponectin also modulates inflammatory processes by modifying expression of inflammatory cytokines, as shown in the model of anti-GBM glomerulonephritis⁵³. Adiponectin deficient mice were more prone to renal injury after administration of anti-GBM serum⁵³.

Plasma adiponectin concentration is almost 3-times higher in haemodialysis patients with chronic kidney disease than in healthy subjects^{54,55}. However increased plasma adiponectin concentration in these patients could not be explained by its oversecretion by adipose tissue, because the expression of the adiponectin gene (ApM1) is decreased in adipocytes of patients with advanced CKD⁵⁶. The kidneys are the

main organ participating in the biodegradation and elimination of adiponectin from the circulation. Thus as expected, successful kidney transplantation is accompanied by prompt reduction, but not normalization, of plasma adiponectin concentration⁵⁷. In addition an inverse relationship between plasma adiponectin concentrations and GFR was found in patients with essential hypertension and apparently healthy individuals^{43,58}. Measurements of plasma adiponectin concentrations in the renal veins and aorta of patients with haemodynamically important renal artery stenosis confirmed the role of the kidneys in the elimination of adiponectin⁵⁹.

Lower adiponectin gene expression in CKD patients may be partially caused by the micro-inflammation. Also in the general population, an inverse relationship was found between concentrations of adiponectin and CRP^{60,61}. The same is true for haemodialysis patients^{55,62,63}. Experimental studies indicating that $\text{TNF}\alpha$ and IL-6 inhibit adiponectin gene expression in cultured adipocytes^{64,65} has already confirmed this clinical observation.

Low plasma adiponectin concentration is now recognised as a new potential risk factor in cardiovascular morbidity and mortality. In a large cohort of hemodialysis patients lower plasma adiponectin concentration was an independent predictor of fatal and non-fatal cardiovascular complications in these patients^{54,55}.

Resistin

Resistin is a large polypeptide which is produced exclusively by adipose tissue matrix⁶⁶. Plasma resistin concentration is markedly elevated in obese overfed insulin-resistant mice and also in ob/ob and db/db mice with obesity and diabetes mellitus⁶⁷. In mice, neutralization of resistin by anti-resistin antibodies lowers glu-

cose concentration and improves insulin sensitivity. Conversely, intraperitoneal administration of resistin causes glucose intolerance and insulin resistance in these animals⁶⁷. Moreover, resistin impairs glucose uptake by adipocytes *in vitro*. It is important to stress that rosiglitazone, an agonist of peroxisome proliferator-activated receptor- γ (PPAR- γ), reduces gene expression and secretion of resistin⁶⁷. In spite of this initial finding in mice, the true role of resistin in humans is unknown. In humans no association between plasma resistin concentration and adiposity or insulin resistance has been found⁶⁸.

Similarly to other adipokines, plasma resistin concentration is elevated with impairment of the kidney function⁶⁹. However, no association between plasma resistin concentration and insulin sensitivity has been found in these patients⁶⁹.

Interleukin-6 (IL-6)

IL-6 is one of the main proinflammatory mediators primarily secreted by the immune system cells⁴. However 20-30% of these cytokines in the circulation is produced by the adipose tissue⁷⁰. In obese subjects with high waist-hip ratio the participation is even greater⁷⁰. It was also shown that omental adipose tissue releases 2-3 times more IL-6 than subcutaneous tissue⁷¹.

The expression of IL-6 gene is related to the adipose cell size⁷² and increases postprandially⁷³. Moreover several studies have documented the positive relationship between BMI and plasma IL-6 concentrations^{74,75}.

In the liver, IL-6 stimulates synthesis of acute-phase response proteins including C-reactive protein (CRP), fibrinogen, serum amyloid-A and α -1 antichymotrypsin⁷⁶. It is already well known that CRP is both a marker and important risk factor of cardiac events and atherosclerosis in the general population and CKD patients⁷⁷. It is

also worth stressing that IL-6 stimulates fibrinogen production and platelets activity, which increases the risk of clot formation⁷⁸.

In CKD patients plasma concentration of IL-6 is higher than in the general population, mainly as a consequence of prolonged plasma high-life of this cytokine and chronic infections such as *Chlamydia pneumonia* and others⁷⁹. Elevated concentration of IL-6 and CRP are both strong predictors of mortality in haemodialysis patients^{80,81}.

Tumour necrosis factor alpha (TNF- α)

TNF- α is predominantly synthesized by macrophages infiltrating adipose tissue^{82,83}. Subcutaneous adipose tissue is claimed to produce more TNF- α than visceral one⁸⁴. This cytokine is involved in the genesis of inflammation and/or insulin resistance⁸⁵.

The excess of TNF- α production by adipose tissue is one of many factors influencing insulin resistance, but certainly not a fundamental one. Mice lacking TNF- α or TNF- α receptor function demonstrate only modest protection against hyperglycemia and insulin resistance when obese^{86,87}.

There are three mechanisms linking TNF- α with insulin resistance. The first is related to inactivation of the insulin receptor signaling pathway (involving NF κ B and or INK) by phosphorylation^{88,89}. The second mechanism engages adiponectin, the secretion of which from adipocytes is markedly reduced by TNF- α ⁴⁶. The third mechanism compromises induction of adipocytes lipolysis and stimulation of hepatic lipogenesis⁹⁰.

The consequence of elevated plasma TNF- α concentration in CKD patients⁹¹ is until now only a matter of speculation.

Plasminogen activator inhibitor-1

Adipose tissue is a site of abundant PAI-1 synthesis⁹². PAI-1 is a pro-coagulative agent and inhibits fibrinolysis. Circulating plasma level of PAI-1 is an independent predictor of coronary artery disease⁹³. These characteristics may explain coronary artery disease in obese patients. Increased PAI concentrations promote release of platelet-derived growth factors which are known to play a role in vascular injury.

The renin-angiotensin system

All components of the renin-angiotensin system are found in adipose tissue^{94,95}. It is still unclear whether angiotensin II secreted by adipose tissue has an important systemic haemodynamic and/or non-haemodynamic effects⁹⁶.

CONCLUSION

Visceral adiposity is the key feature of metabolic syndrome (Figure 1). Adipokines oversecreted by the excessive amount of visceral adipose tissue enables us to understand basic metabolic changes related to obesity, such as insulin resistance, dyslipidaemia and chronic microinflammation. Accelerated atherosclerosis, diabetes type 2 and obesity related hypertension are the most frequent causes of morbidity and mortality of the general population. Chronic kidney diseases are also common consequence of diabetic, hypertensive or ischemic nephropathies. Only in markedly (morbidly) obese humans obesity *per se* may directly impair the kidney function as an isolated entity – obesity related glomerulopathy.

However, even a moderate impairment of kidney function further increases the risk of cardiovascular morbidity and mortality⁹⁷.

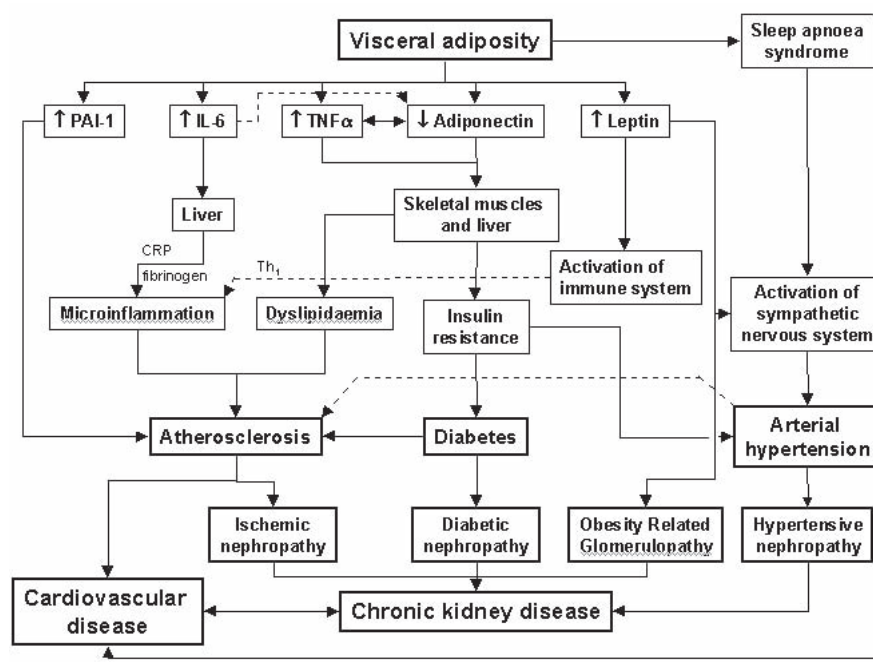


Figure 1 – Visceral adiposity and cardiovascular and kidney diseases.

Therefore it is expected that pharmacological intervention in paracrine/endocrine or immune activity of the adipose tissue may improve the outcome of metabolic syndrome and related, cardiovascular and renal complications in the near future, bringing new hope.

Address for correspondence:

Prof. dr hab. med. Andrzej Więcek, FRCP (Edin)
Department of Nephrology, Endocrinology and Metabolic Diseases,
Silesian University Medical School, Katowice
ul. Francuska 20/24,
40-027 Katowice, POLAND
E-mail: awiecek@spskm.katowice.pl

REFERENCES

1. Havel PJ. Update on adipocyte hormones: regulation of energy balance and carbohydrate/lipid metabolism. *Diabetes* 2004;53:S143.
2. Więcek A, Kokot F, Chudek J, Adamczak M. The adipose tissue - a novel endocrine organ of interest to the nephrologist. *Nephrol Dial Transplant* 2002;17:191-195.
3. Wisse BE. The inflammatory syndrome: the role of adipose tissue cytokines in metabolic disorders linked to obesity. *J Am Soc Nephrol* 2004; 15: 2792-2800.
4. Fain JN, Madan AK, Hiler ML, Cheema P, Bahouth SW. Comparison of the release of adipokines by adipose tissue, adipose tissue matrix, and adipocytes from visceral and subcutaneous abdominal adipose tissues of obese humans. *Endocrinology* 2004; 145: 2273-2282.
5. Löhn M, Dubrovská G, Lauterbach B, Luft FC, Gollasch M, Sharma AM. Periadventitial fat releases a vascular relaxing factor. *FASEB J* 2002; 16: 1057-1063.
6. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM. Positional cloning of the mouse obese gene and its human homologue. *Nature* 1994; 372: 425-432.
7. Muzzin P, Eisensmith RC, Copeland KC, Woo SL. Correction of obesity and diabetes in genetically obese mice by leptin gene therapy. *Proc Natl Acad Sci U S A* 1996; 93: 14804-14808.
8. Horn R, Geldszus R, Potter E, von zur Muhlen A, Brabant G. Radioimmunoassay for the detection of leptin in human serum. *Exp Clin Endocrinol Diabetes* 1996; 104: 454-458.
9. Caro JF, Kolaczynski JW, Nyce MR, et al. Decreased cerebrospinal-fluid/serum leptin ratio in obesity: a possible mechanism for leptin resistance. *Lancet* 1996; 348: 159-161.
10. Flier JS. Physiology: is brain sympathetic to bone? *Nature* 2002; 420: 619, 621-622.
11. Grinspoon S, Gulick T, Askari H, et al. Serum leptin levels in women with anorexia nervosa. *J Clin Endocrinol Metab* 1996; 81: 3861-3863.
12. Ormseth OA, Nicolson M, Pellemounter MA, Boyer BB. Leptin inhibits prehibernation hyperphagia and reduces body weight in arctic ground squirrels. *Am J Physiol* 1996; 271: R1775-R1779.
13. Hwa JJ, Fawzi AB, Graziano MP, et al. Leptin increases energy expenditure and selectively promotes fat metabolism in ob/ob mice. *Am J Physiol* 1997; 272: R1204-R1209.
14. Gibson WT, Farooqi IS, Moreau M, et al. Congenital leptin deficiency due to homozygosity for the Delta133G mutation: report of another case and evaluation of response to four years of leptin therapy. *J Clin Endocrinol Metab* 2004; 89: 4821-4826.
15. Chehab FF, Mounzih K, Lu R, Lim ME. Early onset of reproductive function in normal female mice treated with leptin. *Science* 1997; 275: 88-90.
16. Sierra-Honigmann MR, Nath AK, Murakami C, et al. Biological action of leptin as an angiogenic factor. *Science* 1998; 281: 1683-1686.
17. Lord GM, Matarese G, Howard JK, Baker RJ, Bloom SR, Lechler RI. Leptin modulates the T-cell immune response and reverses starvation-induced immunosuppression. *Nature* 1998; 394: 897-901.
18. Bornstein SR, Licinio J, Tauchnitz R, et al. Plasma leptin levels are increased in survivors of acute sepsis: associated loss of diurnal rhythm, in cortisol and leptin secretion. *J Clin Endocrinol Metab* 1998; 83: 280-283.
19. Hasty AH, Shimano H, Osuga J, et al. Severe hypercholesterolemia, hypertriglyceridemia, and atherosclerosis in mice lacking both leptin and the low density lipoprotein receptor. *J Biol Chem* 2001; 276: 37402-37408.
20. Rahmouni K, Haynes WG, Mark AL. Cardiovascular and sympathetic effects of leptin. *Curr Hypertens Rep* 2002; 4: 119-125.
21. Szanto I, Kahn CR. Selective interaction between leptin and insulin signaling pathways in a hepatic cell line. *Proc Natl Acad Sci USA* 2000; 97: 2355-2360.
22. Adamczak M, Kokot F, Więcek AW. Relationship between plasma renin profile and leptinaemia in patients with essential hypertension. *J Hum Hypertens* 2000; 14: 503-950.
23. Kambham N, Markowitz GS, Valeri AM, Lin JK. Obesity-related glomerulopathy: an emerging epidemic. *Kidney Int* 2001; 59: 1498-1509.
24. Wolf G, Hamann A, Han DC, et al. Leptin stimulates proliferation and TGF-beta expression in renal glomerular endothelial cells: potential role in glomerulosclerosis. *Kidney*

- Int 1999; 56: 860–872.
25. Han DC, Isono M, Chen S, et al. Leptin stimulates type I collagen production in db/db mesangial cells. Glucose uptake and TGF- β type II receptor expression. *Kidney Int* 2001; 59: 1315–1323.
26. Howard JK, Lord GM, Clutterbuck EJ, et al. Plasma immunoreactive leptin concentration in endstage renal disease. *Clin Sci* 1997; 93: 119–126.
27. Heimbürger O, Lönnqvist F, Danielsson A, Nordenström J, Stenvinkel P. Serum immunoreactive leptin concentration and its relation to the body fat content in chronic renal failure. *J Am Soc Nephrol* 1997; 8: 1423–1430.
28. Kokot F, Chudek J, Karkoszka H, Adamczak M, Więcek A, Klimek D. Does PTH influence leptin concentration in haemodialysed uraemic patients? *Nephron* 1999; 82: 372–373.
29. Nordfors L, Lönnqvist F, Heimbürger O, Danielsson A, Schalling M, Stenvinkel P. Low leptin gene expression and hyperleptinemia in chronic renal failure. *Kidney Int* 1998; 54: 1267–1275.
30. Cumin F, Baum HP, Levens N. Mechanism of leptin removal from the circulation by the kidney. *J Endocrinol* 1997; 155: 577–585.
31. Kokot F, Adamczak M, Więcek A. Plasma leptin concentration in kidney transplant patients during the early post-transplant period. *Nephrol Dial Transplant* 1998; 13: 2276–2280.
32. Cumin F, Baum HP, de Gasparo M, Levens N. Removal of endogenous leptin from the circulation by the kidney. *Int J Obes Relat Metab Disord* 1997; 21: 495–504.
33. Ficek R, Kokot F, Chudek J, Adamczak M, Ficek J, Więcek A. Plasma leptin concentration in patients with acute renal failure. *Clin. Nephrol* 2004; 62: 84–91.
34. Zeng J, Patterson BW, Klein S, et al. Whole body leptin kinetics and renal metabolism in vivo. *Am J Physiol* 1997; 273: E1102–E1106.
35. Cheung W, Yu PX, Little BM, Cone RD, Marks DL, Mak RH. Role of leptin and melanocortin signaling in uremia-associated cachexia. *J Clin Invest* 2005; 115: 1659–1665.
36. Chudek J, Adamczak M, Kania M, et al. Does plasma leptin concentration predict the nutritional status of hemodialyzed patients with chronic renal failure? *Med Sci Monit* 2003; 9: CR377–CR382.
37. Merabet E, Dagogo-Jack S, Coyne DW, et al. Increased plasma leptin concentration in end-stage renal disease. *J Clin Endocrinol Metab* 1997; 82: 847–850.
38. Bossola M, Muscaritoli M, Valenza V, et al. Anorexia and serum leptin levels in hemodialysis patients. *Nephron Clin Pract* 2004; 97: c76–c82.
39. Yokota T, Oritani K, Ishikawa JT, et al. Adiponectin, a new member of the family of soluble defense collagens, negatively regulates the growth of myelomonocytic progenitors and the function of macrophages. *Blood* 2000; 96: 1723–1732.
40. Arita Y, Kihara S, Ouchi N, et al. Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun* 1999; 257: 79–83.
41. Hotto K, Funahashi T, Arita Y, et al. Plasma concentration of a novel, adipose-specific protein, adiponectin, in type 2 diabetic patients. *Arterioscler Thromb Vasc Biol* 2000; 20: 1595–1599.
42. Dzielińska Z, Januszewicz A, Więcek A, et al.: Decreased plasma concentration of a novel anti-inflammatory protein-adiponectin-in hypertensive men with coronary artery disease. *Thromb Res* 2003; 110: 365–369.
43. Adamczak M, Więcek A, Funahashi T, Chudek J, Kokot F, Matsuzawa Y. Decreased plasma adiponectin concentration in patients with essential hypertension. *Am J Hypertens* 2003; 16: 72–75.
44. Yamauchi T, Kamon J, Ito Y, et al. Cloning of adiponectin receptors that mediate antidiabetic metabolic effects. *Nature* 2003; 423: 762–769.
45. Ouchi N, Kihara S, Arita Y, et al. Novel modulator for endothelial adhesion molecules adipocyte-derived plasma protein adiponectin. *Circulation* 1999; 100: 2473–2476.
46. Ouchi N, Kihara S, Arita Y et al. Adiponectin and adipocyte-derived plasma protein, inhibits endothelial NF-kappaB signaling through a cAMP-dependent pathway. *Circulation* 2000; 102: 1296–1301.
47. Ouchi N, Kihara S, Arita Y, et al. Adipocyte – derived plasma protein, adiponectin, suppress lipid accumulation and class A receptor expression in human monocyte-derived macrophages. *Circulation* 2001; 103: 1057–1063.
48. Motoshima H, Wu X, Mahadev K, Goldstein BJ. Adiponectin suppresses proliferation and superoxide generation and enhances eNOS activity in endothelial cells treated with oxidized LDL. *Biochem Biophys Res Commun* 2004; 315: 264–271.
49. Okamoto Y, Kihara S, Ouchi N, et al. Adiponectin reduces atherosclerosis in apolipoprotein E-deficient mice. *Circulation* 2002; 106: 2767–2770.
50. Kumada M, Kihara S, Ouchi N, et al. Adiponectin specifically increased tissue inhibitor of metalloproteinase-1 through interleukin-10 expression in human macrophages. *Circulation* 2004; 109: 2046–2049.
51. Yamauchi T, Kamon J, Waki H, et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipodystrophy and obesity. *Nat Med* 2001; 7: 941–946.
52. Spranger J, Kroke A, Mohlig M, et al. Adiponectin and protection against type 2 diabetes mellitus. *Lancet*. 2003; 361: 226–228.
53. Li P, Wu W, Gersch CM, Feng L. Anti-inflammatory effects

- of adiponectin on anti-GBM glomerulonephritis [abstract]. *J Am Soc Nephrol* 2004; 15: 9A.
54. Zoccali C, Mallamaci F, Tripepi G, et al. Adiponectin, metabolic risk factors, and cardiovascular events among patients with end-stage renal disease. *J Am Soc Nephrol* 2002;13:134-141.
55. Ignacy W, Chudek J, Adamczak M, et al. Reciprocal association of plasma adiponectin and serum C-reactive protein concentration in haemodialysis patients with end-stage kidney disease - a follow-up study. *Nephron Clin Pract* 2005; 101: c18-c24.
56. Marchlewska A, Stenvinkel P, Lindholm B, et al. Reduced gene expression of adiponectin in fat tissue from patients with end-stage renal disease. *Kidney Int* 2004; 66: 46-50.
57. Chudek J, Adamczak M, Karkoszka H, et al. Plasma adiponectin concentration before and after successful kidney transplantation. *Transplant Proc* 2003; 35: 2186-2189.
58. Mallamaci F, Zoccali C, Cuzzola F, et al. Adiponectin in essential hypertension. *J Nephrol* 2002; 15: 507-511.
59. Adamczak M, Czerwienńska B, Chudek J, Więcek A. Role of kidneys in metabolism of adiponectin. studies in patients with unilateral renal artery stenosis. [Abstract] XLII Congress ERA-EDTA, Istanbul 2005.
60. Ouchi N, Kihara S, Funahashi T, et al. Reciprocal association of C-reactive protein with adiponectin in blood stream and adipose tissue. *Circulation* 2003; 107: 671-674.
61. Engeli S, Feldpausch M, Gorzelniak K, et al. Association between adiponectin and mediators of inflammation in obese women. *Diabetes* 2003; 52: 942-947.
62. Huang JW, Yen CJ, Chiang HW, Hung KY, Tsai TJ, Wu KD. Adiponectin in peritoneal dialysis patients: a comparison with hemodialysis patients and subjects with normal renal function. *Am J Kidney Dis* 2004; 43: 1047-1055.
63. Lee CT, Lee CH, Su Y, Chuang YC, Tsai TL, Cheni JB. The relationship between inflammatory markers, leptin and adiponectin in chronic hemodialysis patients. *Int J Artif Organs* 2004; 27: 835-841.
64. Fasshauer M, Kralisch S, Klier M, et al. Adiponectin gene expression and secretion is inhibited by interleukin-6 in 3T3-L1 adipocytes. *Biochem Biophys Res Commun* 2003; 301: 1045-1050.
65. Kappes A, Löffler G.: Influences of ionomycin, dibutyryl-cycloAMP and tumour necrosis factor-alpha on intracellular amount and secretion of apM1 in differentiating primary human preadipocytes. *Horm Metab Res* 2000; 32: 548-554.
66. Fain JN, Cheema PS, Bahouth SW, Lloyd Hiler M. Resistin release by human adipose tissue explants in primary culture. *Biochem Biophys Res Commun* 2003; 300: 674-678.
67. Steppan CM, Bailey ST, Bhat S, et al. The hormone resistin links obesity to diabetes. *Nature* 2001; 18: 307-312.
68. Lee JH, Chan JL, Yiannakouris N, et al. Circulating resistin levels are not associated with obesity or insulin resistance in humans and are not regulated by fasting or leptin administration: cross-sectional and interventional studies in normal, insulin-resistant, and diabetic subjects. *J Clin Endocrinol Metab* 2003; 88: 4848-4856.
69. Kielstein JT, Becker B, Graf S, Brabant G, Haller H, Fliser D. Increased resistin blood levels are not associated with insulin resistance in patients with renal disease. *Am J Kidney Dis* 2003; 42: 62-66.
70. Mohamed-Ali V, Goodrick S, Rawesh A, et al. Subcutaneous adipose tissue releases interleukin-6, but not tumor necrosis factor-alpha, in vivo. *J Clin Endocrinol Metab* 1997; 82: 4196-4200.
71. Fried SK, Bunkin DA, Greenberg AS. Omental and subcutaneous adipose tissues of obese subjects release interleukin-6: depot difference and regulation by glucocorticoid. *J Clin Endocrinol Metab* 1998; 83: 847-850.
72. Sopasakis VR, Sandqvist M, Gustafson B, et al. High local concentrations and effects on differentiation implicate interleukin-6 as a paracrine regulator. *Obes Res* 2004; 12: 454-460.
73. Orban Z, Remaley AT, Sampson M, Trajanoski Z, Chrousos GP. The differential effect of food intake and beta-adrenergic stimulation on adipose-derived hormones and cytokines in man. *J Clin Endocrinol Metab* 1999; 84: 2126-2133.
74. Park HS, Park JY, Yu R. Relationship of obesity and visceral adiposity with serum concentrations of CRP, TNF-alpha and IL-6. *Diabetes Res Clin Pract* 2005; 69: 29-35.
75. Maachi M, Pieroni L, Bruckert E, et al. Systemic low-grade inflammation is related to both circulating and adipose tissue TNFalpha, leptin and IL-6 levels in obese women. *Int J Obes Relat Metab Disord* 2004; 28: 993-997.
76. Zoccali C, Mallamaci F, Tripepi G. Adipose tissue as a source of inflammatory cytokines in health and disease: focus on end-stage renal disease. *Kidney Int* 2003; 84 [Suppl]: S65-S68.
77. Burke AP, Tracy RP, Kolodgie F, et al. Elevated C-reactive protein values and atherosclerosis in sudden coronary death: association with different pathologies. *Circulation* 2002; 105: 2019-2023.
78. Burstein SA, Peng J, Friesse P, et al. Cytokine-induced alteration of platelet and hemostatic function. *Stem Cells* 1996; 14 [Suppl 1]: 154-162.
79. Jacobs P, Glorieux G, Vanholder R. Interleukin/cytokine profiles in haemodialysis and in continuous peritoneal dialysis. *Nephrol Dial Transplant* 2004; 19 [Suppl 5]: v41-v45.
80. Kimmel PL, Phillips TM, Simmens SJ, et al. Immunologic function and survival in hemodialysis patients. *Kidney Int* 1998; 54: 236-244.

81. Stenvinkel P, Wanner C, Metzger T, et al. Inflammation and outcome in end-stage renal failure: does female gender constitute a survival advantage? *Kidney Int* 2002; 62: 1791-1798.
82. Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL, Ferrante AW Jr. Obesity is associated with macrophage accumulation in adipose tissue. *J Clin Invest* 2003; 112: 1796-1808.
83. Fain JN, Bahouth SW, Madan AK. TNF α release by the nonfat cells of human adipose tissue. *Int J Obes Relat Metab Disord* 2004; 28: 616-622.
84. Winkler G, Kiss S, Keszthelyi L, et al. Expression of tumor necrosis factor (TNF)- α protein in the subcutaneous and visceral adipose tissue in correlation with adipocyte cell volume, serum TNF- α , soluble serum TNF-receptor-2 concentrations and C-peptide level. *Eur J Endocrinol* 2003; 149: 129-135.
85. Sethi J, Hotamisligil GS. The role of TNF- α in adipocyte metabolism. *Semin Cell Dev Biol* 1999; 10: 19-29.
86. Uysal KT, Wiesbrock SM, Marino MW, Hotamisligil GS. Protection from obesity-induced insulin resistance in mice lacking TNF- α function. *Nature* 1997; 389: 610-614.
87. Ventre J, Doebber T, Wu M, et al. Targeted disruption of the tumor necrosis factor- α gene: metabolic consequences in obese and nonobese mice. *Diabetes* 1997; 46: 1526-1531.
88. Hirosumi J, Tuncman G, Chang, et al. A central role for JNK in obesity and insulin resistance. *Nature* 2002; 420: 333-336.
89. Hotamisligil GS. Inflammatory pathways and insulin action. *Int J Obes Relat Metab Disord* 2003; 27 [Suppl 3]: S53-S55.
90. Feingold KR, Grunfeld C. Role of cytokines in inducing hyperlipidemia. *Diabetes* 1992; 41 [Suppl 2]: 97-101.
91. Nishimura M, Hashimoto T, Kobayashi H, et al. Possible involvement of TNF- α in left ventricular remodeling in hemodialysis patients. *J Nephrol* 2003; 16: 641-649.
92. Alessi MC, Peiretti F, Morange P, Henry M, Nalbone G, Juhan-Vague I. Production of plasminogen activator inhibitor-1 by human adipose tissue: possible link between visceral fat accumulation and vascular disease. *Diabetes* 1997; 46: 860-867.
93. Segarra A, Chacon P, Martinez-Eyarre C, et al. Circulating levels of plasminogen activator inhibitor type-1, tissue plasminogen activator, and thrombomodulin in hemodialysis patients: biochemical correlations and role as independent predictors of coronary artery stenosis. *J Am Soc Nephrol* 2001; 12: 1255-1263.
94. Gorzelniak K, Engeli S, Janke J, Luft FC, Sharma AM. Hormonal regulation of the human adipose-tissue renin-angiotensin system: relationship to obesity and hypertension. *J Hypertens* 2002; 20: 965-973.
95. Karlsson C, Lindell K, Ottosson M, Sjöström L, Carlsson B, Carlsson LM. Human adipose tissue expresses angiotensinogen and enzymes required for its conversion to angiotensin II. *J Clin Endocrinol Metab* 1998; 83: 3925-3929.
96. Klahr S, Morrissey J. Angiotensin II and gene expression in the kidney. *Am J Kidney Dis* 1998; 31: 171-176.
97. Go AS, Chertow GM, Fan D, McCulloh CE, Hsu C. Chronic kidney disease and the risk of death, cardiovascular events, and hospitalization. *N Engl J Med* 2004; 351: 1296-1305.